

REVIEW

Open Access

The amygdala between sensation and affect: a role in pain

Pierre Veinante^{1,2*}, Ipek Yalcin^{1,2} and Michel Barrot^{1,2}

Abstract

The amygdala is a structure of the temporal lobe thought to be involved in assigning emotional significance to environmental information and triggering adapted physiological, behavioral and affective responses. A large body of literature in animals and human implicates the amygdala in fear. Pain having a strong affective and emotional dimension, the amygdala, especially its central nucleus (CeA), has also emerged in the last twenty years as key element of the pain matrix. The CeA receives multiple nociceptive information from the brainstem, as well as highly processed polymodal information from the thalamus and the cerebral cortex. It also possesses the connections that allow influencing most of the descending pain control systems as well as higher centers involved in emotional, affective and cognitive functions. Preclinical studies indicate that the integration of nociceptive inputs in the CeA only marginally contributes to sensory-discriminative components of pain, but rather contributes to associated behavior and affective responses. The CeA doesn't have a major influence on responses to acute nociception in basal condition, but it induces hypoalgesia during aversive situation, such as stress or fear. On the contrary, during persistent pain states (inflammatory, visceral, neuropathic), a long-lasting functional plasticity of CeA activity contributes to an enhancement of the pain experience, including hyperalgesia, aversive behavioral reactions and affective anxiety-like states.

Keywords: Amygdala, Hypoalgesia, Hyperalgesia, Affective/emotional component, Persistent pain

Background

Many cerebral structures, constituting the pain matrix, participate in nociception and in the expression and modulation of pain. They transmit and decode nociceptive information, generate, amplify or reduce the pain sensation, allow expression of defensive or distress behavior, and influence the affect. Among these brain structures, the amygdaloid complex, or amygdala, appears as a key component of the pain matrix [1]. Indeed, while this group of sub-cortical nuclei within the temporal lobe is well known for its critical role in the control of emotions [2], it has also been the target for a large number of fundamental research in the pain field during the two past decades. Overall, the amygdala is considered to provide an emotional value – either positive or negative – to sensory information, thus leading

to adapted behavioral and affective responses and contributing to emotional memory. This role has been more particularly studied in the conditioned fear experimental paradigm [2-4]. Such conditioning is an adaptive process in response to potential danger. As nociceptive stimulations are also potentially harmful for body integrity, it is not surprising that the amygdala can integrate various nociceptive information to initiate or modulate the autonomic and behavioral responses according to the environmental context (internal and external) and to the affective state. This adaptation includes mechanisms of hypoalgesia/analgesia that allows reflex inhibition and facilitates fight or flight responses. Moreover, this brain structure participates in the anticipation/prediction of potential danger, based on salient sensory cues (sounds, images, odors...), to initiate the same adaptive responses as in direct presence of the danger. This is for example leading to conditioned or to stress-induced analgesia.

Pain includes a strong emotional component making it aversive and a reciprocal relation exists between pain and affective states. Stress can either inhibit or amplify

* Correspondence: veinantep@inci-cnrs.unistra.fr

¹Institut des Neurosciences Cellulaires et Intégratives, UPR3212, Centre National de la Recherche Scientifique, 21 Rue René Descartes, 67084, Strasbourg Cedex, France

²Université de Strasbourg, 21 Rue René Descartes, 67084, Strasbourg Cedex, France

pain; and anxiety as well as depressive states are often associated with more intense pain sensation, particularly when pain becomes chronic. In such chronic pain state, the amygdala could contribute to the hyperalgesia as well as to the anxio-depressive consequences of pain.

A few publications reviewed the role of the amygdala in persistent pain [5,6]. The present review will consider the recent published data on the subject and address the key role of amygdala, especially its central nucleus, in various pain processes, either physiological or pathological. This will be done by reminding the anatomical context and presenting the morphological and functional evidences allowing insertion of the amygdala within the pain matrix. Then, we will show that the amygdala can exert either inhibitory or facilitating action on nociception and pain as well as on its affective component and consequences. Finally, we will address recent electrophysiological, neurochemical and biochemical data that are giving insights into underlying cellular and molecular mechanisms. While experimental data that are presented concern studies conducted in rodents, we will confront them with human brain imaging studies, thus stressing the potential role of the amygdala in human pain, especially with clinical relevance.

Review

Amygdala and extended amygdala

The amygdala is a nuclear complex with almond shape, located in the temporal lobe of mammals and identified by Burdach at the start of 19th century. It is an heterogeneous structure, grouping around a dozen of nuclei depending on the considered nomenclature, these nuclei being classically clustered in 4 groups: superficial, basolateral, central and medial (Figure 1) [7,8]. The first two groups constitute the corticobasal amygdala that shows cortical-like characteristics [8]. Its main nuclei, the lateral and the basolateral nuclei are known as the basolateral amygdala (BLA) and mostly contain projection neurons of pyramidal type, synthesizing glutamate. It is reciprocally connected with the cerebral cortex, and sends a major projection to the striatal complex, especially in its ventral part [9,10]. The BLA also densely innervates the central and medial groups of amygdala nuclei. These last two groups display an organization that is similar to the one of the basal ganglia, with striatopallidal-like neuronal morphology [8,11]. Moreover, the central and the medial amygdala groups receive information from the cerebral cortex but do not send any direct projection to it. In addition to the four groups, a few nuclei of the amygdala remain unclassified, among them the intercalated cell masses which are small clusters of densely packed GABAergic neurons [12].

The central nucleus of the amygdala (CeA), main component of the central group of amygdala nuclei, is

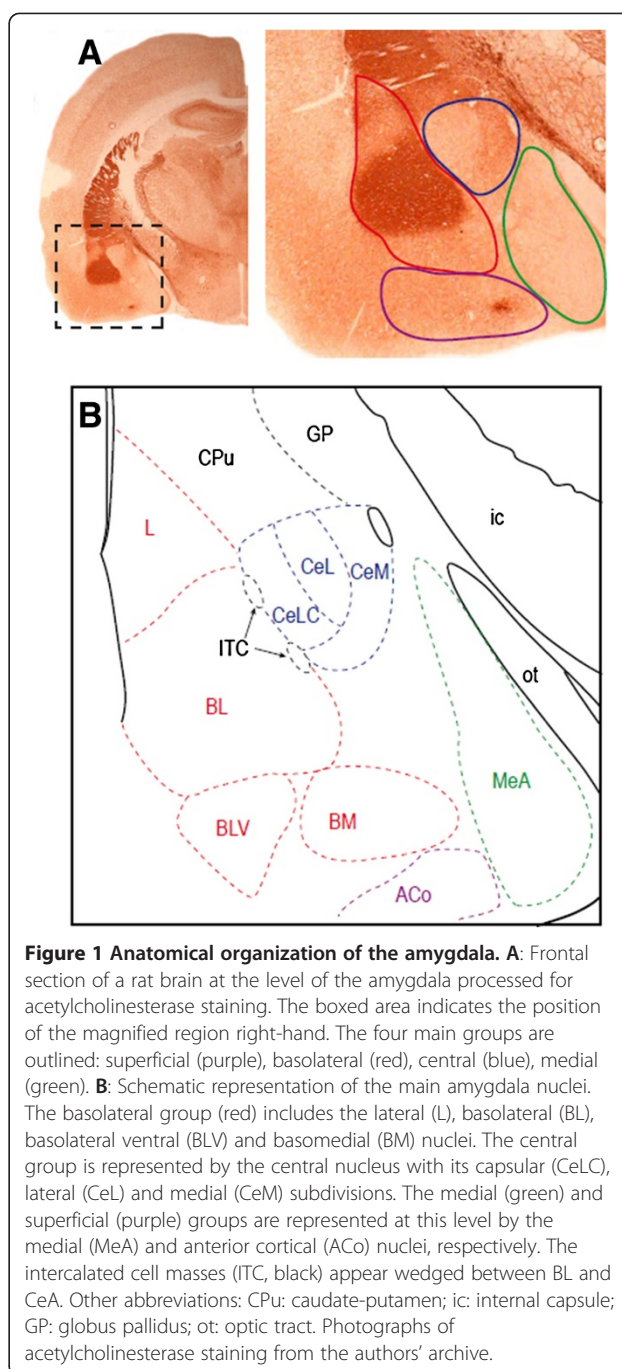


Figure 1 Anatomical organization of the amygdala. **A:** Frontal section of a rat brain at the level of the amygdala processed for acetylcholinesterase staining. The boxed area indicates the position of the magnified region right-hand. The four main groups are outlined: superficial (purple), basolateral (red), central (blue), medial (green). **B:** Schematic representation of the main amygdala nuclei. The basolateral group (red) includes the lateral (L), basolateral (BL), basolateral ventral (BLV) and basomedial (BM) nuclei. The central group is represented by the central nucleus with its capsular (CeLC), lateral (CeL) and medial (CeM) subdivisions. The medial (green) and superficial (purple) groups are represented at this level by the medial (MeA) and anterior cortical (ACo) nuclei, respectively. The intercalated cell masses (ITC, black) appear wedged between BL and CeA. Other abbreviations: CPu: caudate-putamen; ic: internal capsule; GP: globus pallidus; ot: optic tract. Photographs of acetylcholinesterase staining from the authors' archive.

considered as the output nuclei of the amygdaloid complex. It integrates information treated by the corticobasal group and it influences effector centers. The CeA is also reciprocally connected to more rostral forebrain structures, such as the lateral part of the bed nucleus of the stria terminalis (BSTL) and the dorsal part of the substantia innominata. This group of interconnected structures presents strong morphofunctional homogeneity and is designed as the central extended amygdala [8,13]. In this review, we will more specifically detail data

on the CeA, which is the most studied amygdaloid structure for its role in nociception and pain, and also refer to findings on the extended amygdala, as CeA functions are closely related to this forebrain macrostructure.

The CeA has three subdivisions: capsular (CeLC), central (CeL) and medial (CeM), that are defined based on cytoarchitecture, neurochemistry and connectivity [8,11]. The CeLC and the CeL mostly contain medium-size spiny striatal-like neurons, while the CeM main neuronal population has a leptodendritic pallidal-like morphology (i.e. neurons with long dendrites with few or no spines). Neurons from these 3 sub-divisions co-synthesize GABA and neuropeptides, such as opioid peptides (enkephalins, dynorphin, endorphins), corticotropin-releasing factor (CRF), neurotensin, somatostatin, galanin or substance P [11,14-16]. These peptides and others, such as calcitonin gene-related peptide (CGRP), are also present in a dense neuropile of incoming axonal terminals [11]. Connections of the CeA (Figure 2) are characterized by strong relation, often reciprocal, with autonomic and modulatory centers of the hypothalamus and the brainstem. This includes the dorsal vagal complex, the parabrachial nucleus, the periaqueductal gray (PAG), the medullary and mesopontine reticular formation, the paraventricular nucleus of the hypothalamus and the lateral hypothalamus [7,8,11]. The midbrain dopaminergic centers, the locus coeruleus and the dorsal raphe are also under CeA control, and the CeA receives monoaminergic inputs from these structures. The connections of the CeA with the thalamus are sparser but reciprocal, and concern the paraventricular, posterior intralaminar and subparafascicular nuclei [8]. Cortical inputs mainly arise from non-isocortical

areas (prefrontal, insular and perirhinal cortices) and from the corticobasal nuclei of the amygdala [11,17]. While the CeA doesn't directly project to the cerebral cortex, it has outputs toward substantia innominata cholinergic neurons that are projecting to the cortex [8]. The amygdaloid intercalated cell masses also provide an intra-amygdaloid input to the CeA [12]. Last, the CeA is interconnected with other components of the central extended amygdala through a dense network of reciprocal projections. Within the CeA, information follow a latero-medial flux [11]: most afferents do synapses in the CeLC and the CeL, and neurons of these two subdivisions then influence the CeM where most of the efferent projection neurons of the CeA are found (Figure 2). Globally, these morphofunctional characteristics are similar to other components of the central extended amygdala [8]. This general organization of amygdala and extended amygdala described in rodent is similar to the human brain [8,13].

The CeA receives multiple nociceptive informations

Among the various CeA afferents, two main pathways are preferentially conveying nociceptive information (Figure 2). A first pathway originates from the BLA, and conveys highly integrated polymodal information, including nociceptive, from the thalamus and the cerebral cortex, allowing for example fear conditioning [2,4,7]. A cascade of projections, originating in the ventroposterior, posterior, triangular and posterior intralaminar nuclei of the thalamus, the second somatosensory area and the insular cortex, brings nociceptive information to the BLA which in turn transmits it to the CeA [7,17,18]. More precisely, the lateral nucleus of the amygdala projects

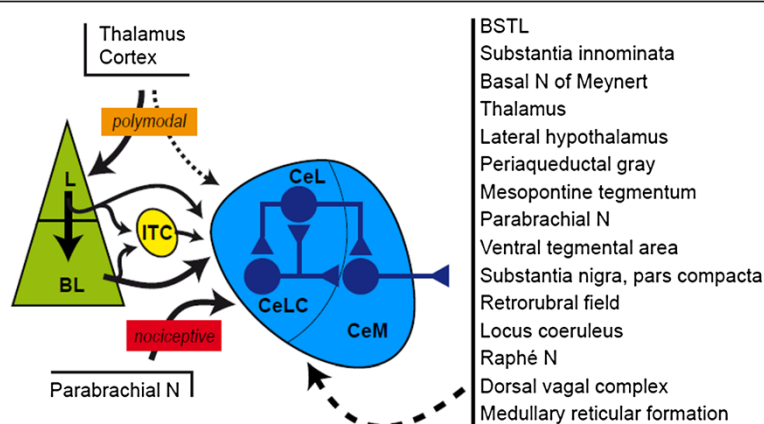


Figure 2 Main connections of the CeA involved in pain processes. The CeA received highly processed polymodal information from the cerebral cortex and the thalamus essentially through inputs from the lateral (L) and basolateral (BL) nuclei, and more direct nociceptive information from the parabrachial nucleus. Intercalated cell masses (ITC) provide an inhibitory input driven by L and BL. Most of these afferents contact neurons in the capsular (CeLC) and lateral (CeL) subdivisions of the central nucleus which control the activity of neurons in the medial subdivision (CeM). Efferents arise mainly from the CeM and target the other components of the central extended amygdala, thalamic nuclei and the integrative, modulator and effector centers in the hypothalamus and the brainstem. Most of these structures send a reciprocal projection to the central nucleus.

lightly to the CeLC and the CeL and massively to the basolateral nucleus which targets preferentially to the CeM [4,7,11]. Additionally, lateral and basolateral projections to the CeA can be relayed in the intercalated cell masses, providing an inhibitory interface in the BLA-CeA pathway [4,12]. Moreover, the CeA also receives less dense but direct projection from these thalamic and cortical areas [17,18]. These largely polysynaptic efferents thus allow the CeA to integrate a nociceptive information which already acquired affective and cognitive significance in cortical circuits.

A second pathway conveys to the CeA more direct and raw nociceptive information. While spinal cord only sparsely sends direct projection to the amygdala, spinal nociceptive information is largely transmitted to the CeA via the parabrachial nucleus (PB). This pontine integrative center is a major target for superficial layers of the spinal cord, but also for deeper layers, for the trigeminal complex and for the nucleus of the solitary tract [19,20]. The nociceptive aspect of these afferents was clearly established by electrophysiology [20]. The PB thus gathers nociceptive information from all cutaneous, deep tissue and visceral territories. Three main ascending pathways are then originating from the PB, toward the medial thalamus, the medial hypothalamus and the central extended amygdala [20]. The parabrachio-amygdaloid pathway is highly organized, with a specific topography connecting various PB subdivisions to the ones of the CeA and the BSTL [19,20]. Moreover, unitary analysis of parabrachio-amygdaloid axonal branching showed that the different components of the central extended amygdala are innervated by axonal collaterals from a same PB neuronal population that never send projection to the medial thalamus or the medial hypothalamus [21].

In vivo electrophysiological studies allowed characterizing the response of CeA neurons to nociceptive stimulations [6,19,20,22,23]. In anesthetized rats, cutaneous (mechanical or thermal), deep tissue (articular and muscular) and visceral nociceptive stimulations provoke changes in the activity of a large majority of CeA neurons (Table 1). Most of them, particularly in the CeLC, are activated by nociceptive stimuli, either exclusively or preferentially, while fewer neuronal inhibition can be observed in the CeL. Few CeM neurons are responding to nociceptive stimuli. This has led to define the CeLC as the “nociceptive amygdala” [6] even though contributions from the CeL to nociceptive process cannot be ruled out. The CeA neurons can encode the stimulus intensity, as shown by sigmoid stimulus–response curves. Receptor fields are particularly large, often bilateral and can cover the whole body [20,22,23]. These neurons often respond to both cutaneous and deep tissue stimulation, whether mechanical or thermal. Together, these data suggest that even though CeA neurons can detect

Table 1 Reported changes in the rodent CeA in different pain situations

Pain situation ^a	Changes in the CeA	Reference
<i>Acute somatic stimulations</i>	Changes (mostly excitation) in electrical activity	[22,23]
<i>Visceral pain</i>		
ip acetic acid	- Increased <i>c-fos</i> mRNA expression	[24]
Esophageal acetic acid	- Increased c-Fos immunoreactivity	[25]
Colitis	- Increased neuronal excitability	[26,27]
	- Enhanced the PB-CeA, but not the BLA-CeA transmission	
	- Increased <i>crf</i> mRNA expression	
Cystitis	- Increased c-Fos and Krox-24 immunoreactivities	[28,29]
	- Increased <i>crf</i> mRNA expression	
<i>Inflammatory pain</i>		
Intraplantar formalin	- Induced ERK activation in the right CeA	[30,31]
Acid-induced muscle pain	- Increased ERK activation	[32]
	- Enhanced the PB-CeA transmission	
Knee joint arthritis	- Increased spontaneous activity in the right CeA	[33-36]
	- Increased neuronal excitability in the right CeA	
	- Enhanced the PB-CeA and the BLA-CeA transmission	
	- Increased mGluR1 and mGluR5 expression	
	- Increased phosphorylation of NR1 subunit	
<i>Neuropathic pain</i>		
Sciatic nerve ligation or section	- Increased spontaneous and evoked activity differentially in the left and right CeA	[37-41]
	- Enhanced the PB-CeA transmission	
	- Increased <i>crf</i> mRNA expression and CRF immunoreactivity	
	- Increased glucocorticoid receptor mRNA expression	
	- Increased cell proliferation	

^a Pain situation: acute somatic stimulations: nociceptive mechanical pinches or thermal stimulations applied on different body parts; ip acetic acid: intraperitoneal injection of acetic acid; colitis: inflammation by intracolonic infusion TNBS (2,4,6-trinitrobenzenesulfonic acid) or zymosan; cystitis: urinary bladder inflammation by intraperitoneal injection of cyclophosphamide; intraplantar formalin: intraplantar injection of diluted formalin in hindpaw; Acid-induced muscle pain: intramuscular injection of acidic saline (pH4) in gastrocnemius; knee-joint arthritis: intraarticular injection of kaolin-caragenan in knee; Sciatic nerve ligation or section: loose or tight ligation of sciatic nerve or spinal roots, or sparing section of roots.

nociceptive information, the stimulus–response shape of this response as well as the wide receptor fields do not allow the CeA to play a major role in the sensoridiscriminative aspect of pain. This brain region rather

contributes to pain controls and to the emotional and affective components of pain [5,6,19,20].

The recruitment of the CeA in visceral and inflammatory pain has also been evidenced using the expression of immediate early genes such as *cfos* (Table 1). For example, intraperitoneal or esophageal acetic acid injection [24,25], colorectal distension [42] or experimental cystitis [28] induce *cFos* in the CeA and the BSTL. Interestingly, this cellular recruitment in the CeA is not observed after intradermal formalin injection in the hindpaw. In this classical inflammatory pain model, *cFos* recruitment rather appears in the basolateral nucleus of the amygdala [24]. However, extracellular signal-regulated kinase (ERK) phosphorylation is observed in the CeA after intraplantar formalin [30] or in acid-induced muscle pain [32]. Last, in visceral pain models but also in more persistent pain models, such as knee intraarticular injection of kaolin-carrageenan, or neuropathic models based on sciatic nerve ligation, major changes can be observed in the CeA (Table 1), such as an increase in spontaneous and evoked activity [33,34,37], an enhancement of synaptic transmission at the PB-CeA and the BLA-CeA synapses [35,38], an increase in the phosphorylation of NR1 glutamatergic subunit [36], an increased expression of CRE, of group I metabotropic glutamatergic receptors (mGluRs) and of corticosterone receptor [26,29,35,39,40] and an increased cell proliferation [41].

Human brain imaging confirms a role of the amygdala in pain processes [1]. However, the spatial resolution only allows a crude discrimination of the various nuclei of the amygdaloid complex. Globally, while older studies failed to detect pain-related changes in the amygdala, more recent studies revealed unilateral and bilateral activation or inhibition. These changes were observed after nociceptive thermal laser stimulation [43,44] or colorectal distension [45]. The amygdala response is usually correlated to the intensity of the nociceptive stimulus, however the importance of attention and emotional factors has also been evidenced. Indeed, repeated thermal nociceptive stimulations of increased intensity lead to an activation of the amygdala that is correlated to the pain perception [43]. However, when sub-threshold stimulus is interpreted as painful by the subject, this situation may activate the amygdala similar to a nociceptive painful stimulus reflecting a state of anxiety or the anticipation of a potentially aversive event. In another study [44], the subjects were informed that they were about to be exposed to a cold painful stimulus lasting 1 minute, or 2 minutes. Functional imaging during the first minute of stimulation revealed that the anticipation of longer stimulus duration led to the deactivation of the amygdala. This suggests that the impact of nociceptive information on the amygdala depends on the context, and not simply of the intrinsic properties of the stimulus.

The CeA influences the nociceptive centers

The CeA receives and integrates nociceptive information, but it also influences the main pain centers. The CeA massively projects to other components of the central extended amygdala, to the lateral hypothalamus and to the brainstem [8]. Anatomical data indicate that the intrinsic and the extrinsic projections to the central extended amygdala arise from distinct neuronal populations [11]. Indeed, studies of unitary axonal reconstruction [46] show that medium size spiny neurons of the CeLC and the CeL mostly project to the BSTL and the dorsal substantia innominata, while axons of CeM projection neurons innervate the various brainstem structure through collateral branches. Since the nociceptive inputs from the PB preferentially target the CeLC and the CeL, such organization suggest that nociceptive information are integrated through an interneuron network intrinsic to the CeA and to the central extended amygdala before being delivered to output neurons influencing effector centers of the brainstem. Some of these centers are part of the nociceptive descending controls. Partially opiodergic CeA outputs are for example innervating the PAG that is a key element of descending controls of nociception through its influence on the ventromedial reticular formation [47]. The PB is also part of descending controls and it receives dense inputs from the CeA [8]. Beside these projections toward integrative centers, the CeA also directly projects to the ventromedial reticular formation [48], the dorsal reticular nucleus [49] and monoaminergic centers such as the substantia nigra and the rostral ventral tegmental area (dopamine), the locus coeruleus (noradrenaline) and the raphe nuclei (serotonin) [48,50-52]. These structures are known to influence the nociceptive message. Moreover, monoaminergic centers projecting to the forebrain modulate striatal and cortical activities, potentially influencing affective, emotional and motivational aspects of pain. Finally, within the central extended amygdala, the dorsal substantia innominata that receives strong inputs from the CeA – and particularly from the CeLC – includes cholinergic neurons that innervate the prefrontal and insular cortices [8,20].

Thus, the CeA has connections allowing the modulation of both the sensory and the affective, emotional and cognitive aspects of pain.

Amygdala, acute pain and analgesia

The potential role of the amygdala in the modulation of pain has been suggested for a long time. It has been clearly shown that the anti- and pro-nociceptive effects are dependent on (1) the type of pain (acute, inflammatory or chronic); (2) the measured parameters (threshold or latency of reflex withdrawal, vocalizations, emotional component); and (3) the emotional state of the subjects (stress, anxiety, fear and expectation).

Nociceptive tests, relying on the latency of appearance of avoidance behavior or on the stimulus threshold, are used to determine the analgesia or hyperalgesia [53]. These tests include for example the tail-flick (tail withdrawal after a thermal stimulus), flinch test (paw or tail withdrawal after electrical stimulation), hot-plate (paw licking or jump after thermal stimulation) or Randall-Selitto (paw withdrawal or vocalization after mechanical stimulation). In these tests, the bilateral lesion or chemical inactivation of the amygdala causes neither antinociceptive nor pro-nociceptive effects in naïve animals [54-56]. However, injections of neuropeptides such as oxytocin, vasopressin, neurotensin, galanin, CRF or CGRP [57-62], or of noradrenergic or cholinergic agonists [63-66] into the CeA induce antinociceptive effects (Table 2). In addition, the amygdala is involved in the analgesic action of systemically administered morphine and cannabinoids since the lesion or the inactivation of the CeA strongly reduces the morphine- or cannabinoid-induced analgesia as measured by the tail-flick test [56,67]. A recent study also showed that deleting the gene of brain-derived neurotrophic factor (BDNF) in parabrachio-amygdaloid neurons decreased the analgesic properties of morphine in mechanical and thermal nociceptive modalities [68]. Finally, the antinociceptive properties of intra-CeA morphine or intra-CeA β -endorphin are dependent upon opiodergic transmission in the amygdala-PAG pathway [47].

These data therefore show that even if the CeA is not directly involved in the modulation of basal nociceptive thresholds in normal situations, it strongly influences analgesia processes (Table 2).

Stressful situations (restraint or noise) and fear, especially the experimental model of fear conditioning, cause analgesia as measured by the tail flick or the hot-plate tests. Several studies demonstrated that bilateral lesion or inactivation of the CeA diminishes or abolishes this analgesic effect [55,69,70,80,92] (Table 2). In parallel, clinical studies showed that stressful stimuli, either painful (electric shocks) or painless (unexpected noises), and innocuous stimuli, previously associated with electric shocks (fear conditioned), cause hypoalgesia in a finger withdrawal test following thermal stimulation [93]. In contrast, anxiety caused by electric shock induces hyperalgesia [93]. Although the direct role of the amygdala in these effects has not yet been shown in humans, the results in rodents are consistent with clearly defined functions of the amygdala in emotional responses to different aversive situations [2].

The amygdala thus can participate in adaptive processes leading to alleviation of pain sensation but its role is not limited to antinociception. While CeA lesion can block the antinociception induced in the tail-flick test by electrical shock pre-exposure, the affective hyperalgesia,

measured by the latency to vocalize after the shock is also reduced by the same CeA lesion [92]. Interestingly, similar results are observed following lesion of the BSTL [92]. Pharmacological manipulation of the CeA can also increase visceromotor and/or somatomotor reflexes in naïve animals, as shown by intra-CeA injection of agonists to group I mGluRs [76,77]. Similarly, CGRP and CRF administration in the CeA can induce mechanical hypersensitivity [82,84], which is in contradiction with older studies reporting antinociceptive effects of these peptides [58,59] (see below). Finally, corticosterone implants in the CeA sensitize the visceromotor reflexes to colorectal and urinary bladder distension [87,88] and increases mechanical sensitivity [89]. The amygdala is thus likely to contribute to both analgesia and hyperalgesia.

Amygdala, sustained pain, hyperalgesia and affective component

Persistent pain, such as inflammatory or neuropathic pain, has a different profile than acute pain. While animal studies often don't address all the symptoms described in humans, the models that are used can display spontaneous nociceptive behaviors, hyperalgesia and/or allodynia, aversion as well as the emotional consequences of pain such as anxiety and depression [94].

The role of the CeA in sustained pain has been examined after intraplantar injection of formalin (somatic inflammatory pain), intraperitoneal injection of acetic acid (visceral inflammatory pain), intraarticular injection of kaolin and carrageenan (somatic inflammatory pain), and ligation/compression of the sciatic nerve (neuropathic pain) (Table 2). Generally, the manipulation of CeA activity, either by activation or by inactivation, did not modify the spontaneous nociceptive behaviors [30,71,72]. However, persistent pain leads to increased neuronal activity and synaptic transmission in the CeA (Table 1) which may be involved in the induction and/or maintenance of hypersensitivity observed in these models [5,6,33,38]. Thus, inhibiting the extracellular signal-regulated kinase (ERK) activation in the CeA decreases mechanical, but not thermal, hyperalgesia in formalin and arthritis models, while the direct activation of ERK in the CeA is sufficient to produce mechanical hyperalgesia in naïve animals [30,91]. In addition, intra-CeA injections of mGluRs antagonist (groups I and III) or of CGRP1 or CRF1 antagonists alleviate the increased withdrawal reflexes as well as audible and ultrasonic vocalizations in monoarthritic animals [78,79,83,85,86]. In neuropathic rats, activation of GABA-A receptors in the CeA also diminishes mechanical hypersensitivity [74]. These data suggest that changes in the activity and neurochemistry of the CeA contribute to the exacerbation of nociceptive responses in persistent pain.

Table 2 Effects of CeA manipulation on pain-related outcomes in different pain models

Pain type ^a	Pain related outcome ^b		Reference
	Nociceptive behavior	Affective/emotional	
1. CeA lesion			
Naïve	- Reduced morphine-induced, stress-induced and conditioned hypoalgesia		[55,56,69]
Formalin	- Reduced morphine-induced and conditioned hypoalgesia	- Decreased pain-induced CPA	[70-73]
Acetic acid		- Decreased pain-induced CPA	[72]
2. Injection of muscimol			
Neuropathy	- Reduced mechanical hyperalgesia	- Decreased escape/avoidance	[74]
3. Injection of NMDA antagonist			
Neuropathy		- Decreased pain-induced CPA	[75]
4. Injection of group I mGluRs ligands			
Naïve	- Agonist induced visceral and mechanical hypersensitivity		[76,77]
	- Antagonist reduced visceral sensitivity		
Formalin	- Antagonist reduced mechanical hypersensitivity		[77]
Arthritis	- Antagonist reduced mechanical hypersensitivity	- Antagonist decreased vocalizations	[78]
Neuropathy		- Agonist increased, and antagonist decreased, pain-induced CPA	[75]
5. Injection of group III mGluRs agonists			
Naïve	- Decreased mechanical sensitivity (mGluR7)	- Decreased vocalizations and anxiety	[79]
Arthritis	- Increased mechanical sensitivity (mGluR8)	- Increased vocalizations and anxiety	[79]
6. Injection of cholinergic agonists			
Naïve	- Decreased thermal sensitivity, reduced jaw opening reflex	- Decreased vocalizations	[63,65,66]
7. Injection of noradrenergic α_2 ligands			
Naïve	- Agonist induced mechanical and thermal hypoalgesia		[64,80]
	- Antagonist reduced stress-induced thermal hypoalgesia		
Acetic acid		- Agonist decreased pain-induced CPA	[81]
8. Injection of noradrenergic β antagonists			
Acetic acid		- Decreased pain-induced CPA	[81]
9. Injection of CGRP receptor ligands			
Naïve	- CGRP decreased mechanical and thermal reflexes		[59]
Naïve	- CGRP increased mechanical reflexes	- CGRP increased vocalizations	[82]
Arthritis	- CGRP1 antagonist inhibited the enhanced reflex to mechanical stimulus	- CGRP1 antagonist decreased vocalizations	[83]
10. Injection of CRF receptor ligands			
Naïve	- CRF decreased mechanical and thermal sensitivity		[58]
Naïve	- CRF increases mechanical sensitivity	- CRF increased vocalizations	[84]
Arthritis	- CRF1 antagonist reduced mechanical hypersensitivity	- CRF1 antagonist decreased vocalizations and anxiety	[85,86]
11. Injection of oxytocin, vasopressin, neurotensin, galanin			
Naïve	- Decreased mechanical and/or thermal sensitivity		[57,60-62]
12. Injection of opioid receptors ligands			
Naïve	- Morphine and β -endorphin induced mechanical and thermal hypoalgesia	- Morphine decreased vocalizations	[47,66]
13. Corticosterone implants			
Naïve	- Sensitized visceromotor reflexes to colorectal and urinary bladder distension and to somatic mechanical sensitivity	- Increased anxiety	[87-90]

Table 2 Effects of CeA manipulation on pain-related outcomes in different pain models (Continued)

<i>14. BDNF gene deletion in the PB-CeA pathway</i>			
Naïve	- Decreased morphine-induced mechanical and thermal hypoalgesia		[68]
<i>15. Intracellular effectors</i>			
Naïve	- ERK activator induced mechanical hypersensitivity		[30]
Formalin	- ERK activation inhibitor decreased mechanical hypersensitivity		[30,31]
Arthritis	- ERK activation inhibitor and PKA inhibitor decreased mechanical hypersensitivity	- ERK activation inhibitor and PKA inhibitor decreased vocalizations	[91]

^aPain type. naïve nociceptive testing on naïve animals; formalin: intraplantar injection of diluted formalin in hindpaw; acetic: intraperitoneal injection of acetic acid; arthritis: intraarticular injection of kaolin-caragenean in knee; neuropathy: compression or ligation of sciatic nerve or spinal root.

^bPain related outcome. thermal sensitivity: latency of withdrawal or escape in tail-flick, hot-plate or Hargreaves tests; mechanical sensitivity: latency or threshold to withdrawal in von Frey or Randall-Sellito tests; CPA: pain-induced conditioned place aversion; vocalizations: intensity/duration/threshold of vocalizations to electrical shock (naïve) or mechanical compression of knee (naïve, arthritis); anxiety: anxiety-like behavior in elevated plus maze test.

The affective and emotional dimensions of pain can also be under the influence of the amygdala. The effects of pharmacological manipulations of the CeA on vocalizations reported above (see also Table 1) are pertinent in this context. Indeed, while audible vocalizations induced by a nociceptive stimulus are believed to reflect nocifensive responses, ultrasonic vocalizations would reflect an affective pain response [78,95]. A widely used test to assess the affective/emotional component of pain, is the pain-induced conditioned place aversion. This behavior is suppressed in the intraplantar formalin and intraperitoneal acetic acid models by bilateral lesions of the CeA [72,73] or by injection of β -adrenoceptor antagonist [81]. Similarly, intra-CeA administration of a GABA-A receptor agonist, a NMDA antagonist or a group I mGluR antagonist reduces the place avoidance behavior in neuropathic rats [74,75].

Finally, the CeA may be a part of the mechanism linking sustained pain to anxiety and depression-like states. It has been shown that the models of monoarthritic pain or of sciatic nerve ligation induce anxiety that is correlated with the activation of the CRF system in the CeA. The expression of this neuropeptide, which has a well established role in anxiety, is increased in the CeA of neuropathic animals [39,40], while the intra-CeA injection of CRF1 antagonist reduces anxious behaviors produced by knee monoarthritis [85,86]. In addition, the pronociceptive effects of corticosterone implants in the CeA are accompanied by anxiety-like behaviors [90].

In humans, brain imaging studies [1] demonstrated changes in the activity of the amygdala of patients with irritable bowel syndrome [45], arthritis [96] or mononeuropathy [97], suggesting that the amygdala may be involved in the emotional aspects related to these pathologies. Interestingly, a brain imaging study showed the participation of an amygdala - anterior cingulate cortex circuit in the higher subjective perception of pain in healthy subjects experiencing sadness [98]. Conversely, viewing pictures of a romantic

partner reduced self-reported pain, in association with activation of the amygdala [99]. These data reinforce the role of amygdala in emotional impact on pain.

In summary, the amygdala, particularly the CeA, may have a mainly antinociceptive influence in acute/phasic pain conditions associated with situations of stress or fear, and a mainly pronociceptive influence in persistent/tonic pain conditions which concerns the sensory as well as the emotional and affective dimensions.

Potential mechanisms with several actors

Some information is available on the cellular and molecular mechanisms by which the amygdala can exert its bidirectional effects on pain parameters. Projections from the PB to the CeA give direct information about the perceived nociceptive stimulation through glutamatergic and peptidergic synapses [100-102], while the glutamatergic projections from the BLA to the CeA can bring polymodal information with an affective valence [5]. This latter input can be turned into an inhibitory control through the intercalated cell masses, functionally placed between the BLA and the CeA [4,12]. The influence of the CeA on its targets will depend on the confrontation of these inputs with other afferent information. The internal network of the CeA and of the central extended amygdala is based on GABAergic inhibitory neurons [103], and the output of this system can either be inhibitory or disinhibitory on target structures [4,104,105]. Changes in the level of pain perception seem to involve the projection of the CeA to the centers of descending pain control, especially the PAG and the ventromedial reticular formation [47], while the influence of the amygdala on the emotional and affective dimensions of pain arises from a larger network that involves indirect connections with cortical regions such as the insular and the cingulate cortex [5,20].

It is now clearly established that persistent pain causes long-lasting changes in the activity of the CeA that could account for its pronociceptive influence on sensory and affective components of pain [5,6]. In arthritic, visceral

and neuropathic pain models, subpopulations of nociceptive neurons of the CeA exhibit increased membrane excitability, leading to higher spontaneous activity, as well as potentiation of synaptic transmission [27,33,35,37,38]. This synaptic plasticity, observed at both the PB-CeA and the BLA-CeA synapses can potentiate nociceptive transmission. Some specificity can be observed concerning pain-related plasticity in the CeA. Indeed, in arthritis model, the population of multireceptive neurons, responding to innocuous stimuli but preferentially to nociceptive stimuli, presents increased basal activity and increased responses to mechanical but not thermal nociceptive stimuli. A second neuronal population, normally insensitive to somatosensory stimulation, develops responses to mechanical, but not thermal nociceptive stimuli. In contrast, arthritis modifies neither the basal activity nor the responses of the nociceptive-specific neurons [33]. In addition, while the PB and the BLA can synapse onto the same neurons, but on different dendritic compartments [106], specific plastic changes can occur with different pain models. While arthritic and neuropathic pain models potentiate both the PB-CeA and the BLA-CeA synapses [33,38], a model of visceral pain only potentiates the PB-CeA synapses [27].

An intriguing observation is that the right CeA appears largely more implicated in persistent pain than the left CeA. Indeed, the activation of ERK after intraplantar formalin is restricted to the right CeA, and the blockade of ERK activation in the right CeA decreased mechanical hyperalgesia at both hindpaws, irrespective to the side of formalin injection [30,31]. In the arthritis model, the enhanced background activity and evoked responses are observed only in the right CeA, as well as the decreased activity following injection of a PKA inhibitor [34]. Finally, in neuropathic rats, spontaneous activity and evoked responses increased in the left CeA 2 and 6 days after sciatic nerve ligation, but declined afterward, whereas these electrophysiological parameters increased in the right CeA at 14 days post-ligation [37]. The different implication of left and right CeA in pain processes could partially account for the conflicting reports indicating antinociceptive [58,59] and pronociceptive [82,84] effects of CRF and CGRP injected into the CeA of naive rats. The anatomical and functional basis of this lateralization remains unclear, but it suggests a strong relation between right amygdala and affective/emotional component of persistent pain.

A number of neurochemical and molecular mechanisms involved in the sensitization of the CeA have been elucidated. They especially implicate the modification of pre- and post-synaptic glutamatergic transmission, with an impact on GABAergic transmission, along with neuropeptidergic modulation [5,6] (Figure 3). Following induction of experimental arthritis, the pre-synaptic expression of mGluR1 (group I) is increased, which in turn

potentiates the synaptic transmission [35,107]. This effect seems to involve a decreased inhibition, resulting in disinhibition of CeLC neurons and of glutamatergic terminals [108]. On the other hand, the activation of group II and III pre-synaptic mGluRs diminishes synaptic transmission [109-111]. At post-synaptic level, activation of group I mGluR (mGluR5) in the CeLC increased the excitatory response to nociceptive input in normal condition but not in arthritic condition [108], possibly involving PKA and ERK activation through reactive oxygen species [112]. In the same inflammatory pain model, NMDA ionotropic receptors are implicated in post-synaptic sensitization, via a PKA-dependant mechanism [36,113], while a NMDA-independent synaptic plasticity is induced in a neuropathic pain model [38]. Last, neuropeptide receptors (CRF1, CGRP1) contribute to this plasticity by favoring NMDA receptor phosphorylation through the recruitment of PKA and ERK kinases [82-84]. Together, these mechanisms lead to both increased nociceptive transmission and neuron sensitization in the CeA. The implication of CRF is here important as it also plays a role in stress response and anxiety. Moreover, while CGRP and CRF in CeA have a pronociceptive influence in arthritis model, the same neuropeptides can be acutely antinociceptive when directly injected in the CeA of control animals [58,59]. This dual action, which likely implies qualitative and quantitative receptor changes, may also exist for other neuropeptides present in CeA, such as oxytocin or neurotensin. Opioid system in the CeA may be important in these phenomena, but its contributions still remain elusive. The three types of opioid receptors (μ , δ , κ) are expressed in the CeA, both pre- and post-synaptically, as well as their endogenous ligands [15,16,114-116]. The opioid system is a key actor in the CeA, due to the enkephalin contribution to intrinsic communications within the central extended amygdala and due to the contribution of the other opioid peptides to extrinsic CeA outputs [11]. The influence of the opioid system in the amygdala on pain is supported by various data, including the antinociceptive action of morphine in the CeA, the synergy between the CeA and the PAG co-delivery of opioid ligands [47] and the changes in CeA opioid receptor binding in neuropathic mice [117]. However, the circuit architecture underlying these actions, and the individual role of each peptide and receptor, is not yet clearly defined. For example, the μ -opioid receptors are pre-synaptically present in the CeA, where they can inhibit glutamate and GABA release from afferents [115,118], but they also act post-synaptically to inhibit CeA interneurons and projection neurons [116] (Figure 3). Thus, these receptors can either inhibit or activate each component of the CeA. It is also not yet known how the opioid system interacts with other neurotransmission systems within the

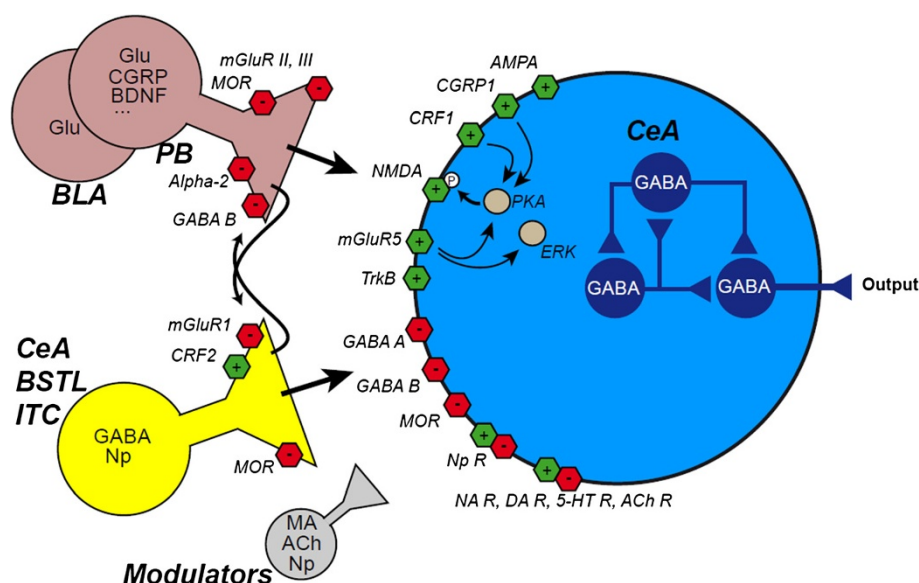


Figure 3 Molecular actors in nociceptive processing in the CeA during arthritic pain. The CeA received nociceptive inputs from glutamatergic (Glu) neurons in the basolateral amygdala (BLA) and in the parabrachial nucleus (PB). GABAergic innervation originates from intrinsic source (CeA and BSTL) and from the intercalated cell masses (ITC). Additional modulation is provided by monoaminergic (MA) and cholinergic (ACh) afferents and by neuropeptides (Np), especially from CeA, BSTL and PB. These neuromediators act on excitatory (green) and inhibitory (red) receptors at pre and post-synaptic levels. In the knee monoarthritis model [5], several mechanisms contribute to an enhanced neurotransmission. At pre-synaptic level, mGluR1 receptors inhibit GABA release, while mGluR from groups II and III decrease glutamate release. At post-synaptic level, mGluR5 receptors are involved in normal transmission in naive animals and NMDA and AMPA ionotropic receptors have also an enhanced activity increasing neuronal sensitivity. CGRP, from PB, and CRF, possibly from the central extended amygdala itself, activate their post-synaptic receptors, CGRP1 and CRF1, respectively. These receptors can activate the protein kinase A (PKA) and thus increase NMDA phosphorylation. The extracellular regulated-signal kinase ERK also contributes to the sensitization of CeA neurons, possibly through a cascade initiated by mGluR5 receptors. CRF can also decrease the activity of CeA neurons, via pre-synaptic CRF2 receptors increasing GABA release. This leads to the inhibition of CeA neurons by activation of post-synaptic GABA A and GABA B receptors and by inhibition of glutamate release by pre-synaptic GABA B receptors. Other neurotransmission system involving noradrenergic (NA, especially through pre-synaptic α_2 -receptors), dopaminergic (DA), serotonergic (5-HT) and cholinergic (ACh) receptors can modulate CeA activity, as well as other neuropeptides, among them opioids acting on pre- and post-synaptic mu receptors (MOR), and BDNF acting on TrkB receptors. Overall, these mechanisms lead to the modulation of the intra-CeA circuitry, based on GABAergic interactions (in blue) and of the CeA output.

CeA, but several studies suggest that the CeA opioid system act downstream to agents that promote antinociception such as galanin or BDNF [60,68]. Finally, adrenoceptors modulate pain processes in the CeA [64,80,81], especially through α_2 adrenoceptors located on PB-CeA pre-synaptic elements which strongly regulate glutamate release [100].

Beyond the central amygdala

While we focused this review on the CeA, it is necessary to remind the contribution of BSTL and BLA to pain process.

The CeA belongs to the central extended amygdala; it is largely interconnected with the BSTL and shares similar afferents, including polymodal and nociceptive inputs from the BLA and the PB [8,21]. While there is no report of an influence of the BSTL on nociceptive sensitivity, a few studies addressed its role in pain-induced conditioned place aversion. Bilateral lesion of the BSTL [119], as well as intra-BSTL injection of a β -adrenoceptor antagonist [120,121], α_2 -adrenoceptor agonist [122] or

CRF1 antagonist [123] decrease place aversion in the intraplantar formalin and/or intraperitoneal acetic acid models. Moreover, noradrenaline and CRF release in the BSTL is enhanced in these pain models [120-122] and CRF mRNA is upregulated in the BSTL of neuropathic rats [40]. These data suggest that the BSTL can be involved in affective/emotional component of pain, maybe in a complementary manner with the CeA, especially in view of BSTL implication in anxiety [3].

The BLA, beyond its role as a mere input provider to the CeA and BSTL, also appears as an important actor in pain processes. Some observations suggest that the BLA and the CeA can have a parallel role in nociceptive responses. Stress- and fear-induced analgesia are reduced by intra-BLA infusion of cannabinoid CB1 receptors antagonists, of muscimol or of diazepam [124-127]; mu opioid agonist, including morphine, injected in the BLA decreases vocalizations to tail shocks [128] and thermal nociceptive sensitivity [129,130]. Sustained pain also impacts the BLA as intraplantar formalin induces *c-fos* mRNA [24], arthritis

increases the expression of the pro-nociceptive cytokine tumor necrosis factor α (TNF- α) [131] and neuropathy increases cell proliferation in the BLA [41]. A functional plasticity of BLA neurons is also observed in arthritis model, characterized by an increased in spontaneous and evoked activity and enhanced synaptic transmission. This pain-induced plasticity can be reversed by CRF1 antagonism [132].

The affective component of pain can also be modulated by the BLA. The formalin-induced conditioned place aversion is reduced by lesion of the BLA [72], as it is the case for the CeA and the BSTL, but also by BLA injection of NMDA antagonist or of morphine [133]. In addition, neutralizing BLA TNF- α with Infliximab antibodies reduced anxiety-like behaviors associated to arthritis [131]. Finally, CRF1 antagonism in BLA of arthritic rats decreased mechanical hypersensitivity, but also pain-related vocalizations and anxiety [132].

These influences of BLA on sensory and affective/emotional aspects of pain can, at least in part, involve its outputs to the CeA and/or BSTL. However, the BLA also appears to be implicated in pain-related cognitive impairment. Indeed, in arthritic rats, blockade of pain-induced plasticity in the BLA with CRF1 antagonism reversed the deficit in decision making evaluated in a gambling task. This deficit has been shown to be dependent upon a deactivation of the medial prefrontal cortex driven by the hyperactivity in the BLA. Importantly, this mechanism appears independent of the CeA [132]. Thus, the connections of the BLA with the cerebral cortex, especially prefrontal, cingulate and insular cortices, could allow the amygdala to influence both the affective/emotional and the cognitive aspects of pain. In addition, the dense BLA input to the ventral striatum [9,10] could also participate in pain-induced changes in motivation and goal-directed behaviors.

Conclusions

The data from the literature show that the amygdala, and in particular the CeA, contributes to pain processes. Its anatomical and functional relations with pain ascending systems, with pain facilitating or inhibitory descending systems, and with affective and cognitive centers are placing the CeA in a critical position within the pain matrix. Acute pain reactions only require CeA if they occur in specific emotional or adaptive contexts. On the other side, persistent and chronic pains alter CeA activity, which influences the pain experience and the related emotional, affective and motivational states. Reciprocally, these states can modify the amygdala processing of information and its impact on pain.

The amygdala has been mostly considered for its roles in emotion, especially in fear [2,4]. While most studies focused on the BLA, recent researches on the fear circuit

disclosed a part of the CeA microcircuitry involved in fear conditioning [104,105]. It is very likely that a similar, if not the same, circuit underlies CeA functions in pain. Indeed, a recent study showed for the first time that the “nociceptive” amygdala (CeLC) undergoes synaptic plasticity at the PB-CeA and the BLA-CeA synapses during fear conditioning [134]. This result emphasizes the need to consider the strong relation between pain and emotion. The contribution to the pain matrix of the CeA, of other components of the amygdala and of the central extended amygdala, should foster researches in the context of chronic pain and the associated anxio-depressive disorders.

Abbreviations

BLA: Basolateral amygdala; CeA: Central amygdaloid nucleus; CeL: Lateral part of the central amygdaloid nucleus; CeLC: Capsular part of the central amygdaloid nucleus; CeM: Medial part of the central amygdaloid nucleus; CGRP: Calcitonin gene-related peptide; CRF: Corticotropin-releasing factor; ERK: Extracellular signal-regulated kinase; mGluR: Metabotropic glutamatergic receptor; PB: Parabrachial nucleus; PKA: Protein kinase A; TNF- α : Tumor necrosis factor α .

Competing interests

Dr. Barrot received lecture fees from Adir and Lilly France, and contract from Missions-Cadres. Drs. Barrot and Yalcin reported a CNRS-filed patent for pain treatments. Other authors reported no potential conflicts of interest.

Authors' contributions

PV conceived this review; PV, IY and MB wrote and revised the manuscript. All authors read and approved the final manuscript.

Acknowledgements

This work was supported by Centre National de la Recherche Scientifique (UPR3212) and University of Strasbourg.

Received: 19 March 2013 Accepted: 11 May 2013

Published: 5 June 2013

References

1. Simons LE, Moulton EA, Linnman C, Carpino E, Becerra L, Borsook D: **The human amygdala and pain: evidence from neuroimaging.** *Hum Brain Mapp* 2012. doi:10.1002/hbm.22199. in press.
2. LeDoux JE: **Emotion circuits in the brain.** *Annu Rev Neurosci* 2000, **23**:155–184.
3. Davis M, Walker DL, Miles L, Grillon C: **Phasic vs sustained fear in rats and humans: role of the extended amygdala in fear vs anxiety.** *Neuropsychopharmacology* 2010, **35**:105–135.
4. Pare D, Duvarci S: **Amygdala microcircuits mediating fear expression and extinction.** *Curr Opin Neurobiol* 2012, **22**:717–723.
5. Neugebauer V, Galhardo V, Maione S, Mackey SC: **Forebrain pain mechanisms.** *Brain Res Rev* 2009, **60**:226–242.
6. Neugebauer V, Li W, Bird GC, Han JS: **The amygdala and persistent pain.** *Neuroscientist* 2004, **10**:221–234.
7. Sah P, Faber ES, Lopez De Armentia M, Power J: **The amygdaloid complex: anatomy and physiology.** *Physiol Rev* 2003, **83**:803–834.
8. Alheid GF, de Olmos JS, Beltramino CA: **Amygdala and extended amygdala.** In *The Rat Nervous System*. 2nd edition. Edited by Paxinos G. San Diego, CA: Academic; 1995:495–578.
9. Buot A, Yelnik J: **Functional anatomy of the basal ganglia: limbic aspects.** *Rev Neurol (Paris)* 2012, **168**:569–575.
10. Fudge JL, Breitbart MA, McClain C: **Amygdaloid inputs define a caudal component of the ventral striatum in primates.** *J Comp Neurol* 2004, **476**:330–347.
11. Cassell MD, Freedman LJ, Shi C: **The intrinsic organization of the central extended amygdala.** *Ann N Y Acad Sci* 1999, **877**:217–241.

12. Palomares-Castillo E, Hernandez-Perez OR, Perez-Carrera D, Crespo-Ramirez M, Fuxe K, Perez de la Mora M: **The intercalated paracapsular islands as a module for integration of signals regulating anxiety in the amygdala.** *Brain Res* 2012, **1476**:211–234.
13. Heimer L, Van Hoesen GW, Trimble M, Zahm DS: *Anatomy of neuropsychiatry: the new anatomy of the basal forebrain and its implications for neuropsychiatric illness.* Amsterdam: Boston Academic Press/Elsevier; 2008.
14. Veinante P, Stoeckel ME, Freund-Mercier MJ: **GABA- and peptide-immunoreactivities co-localize in the rat central extended amygdala.** *NeuroReport* 1997, **8**:2985–2989.
15. Poulin JF, Castonguay-Lebel Z, Laforest S, Drolet G: **Enkephalin co-expression with classic neurotransmitters in the amygdaloid complex of the rat.** *J Comp Neurol* 2008, **506**:943–959.
16. Marchant NJ, Densmore VS, Osborne PB: **Coexpression of prodynorphin and corticotrophin-releasing hormone in the rat central amygdala: evidence of two distinct endogenous opioid systems in the lateral division.** *J Comp Neurol* 2007, **504**:702–715.
17. McDonald AJ, Shammah-Lagnado SJ, Shi C, Davis M: **Cortical afferents to the extended amygdala.** *Ann N Y Acad Sci* 1999, **877**:309–338.
18. Shi C, Davis M: **Pain pathways involved in fear conditioning measured with fear-potentiated startle: lesion studies.** *J Neurosci* 1999, **19**:420–430.
19. Bernard JF, Bester H, Besson JM: **Involvement of the spino-parabrachio-amygdaloid and -hypothalamic pathways in the autonomic and affective emotional aspects of pain.** *Prog Brain Res* 1996, **107**:243–255.
20. Gauriau C, Bernard JF: **Pain pathways and parabrachial circuits in the rat.** *Exp Physiol* 2002, **87**:251–258.
21. Sarhan M, Freund-Mercier MJ, Veinante P: **Branching patterns of parabrachial neurons projecting to the central extended amygdala: single axonal reconstructions.** *J Comp Neurol* 2005, **491**:418–442.
22. Bernard JF, Huang GF, Besson JM: **Nucleus centralis of the amygdala and the globus pallidus ventralis: electrophysiological evidence for an involvement in pain processes.** *J Neurophysiol* 1992, **68**:551–569.
23. Neugebauer V, Li W: **Processing of nociceptive mechanical and thermal information in central amygdala neurons with knee-joint input.** *J Neurophysiol* 2002, **87**:103–112.
24. Nakagawa T, Katsuya A, Tanimoto S, Yamamoto J, Yamauchi Y, Minami M, Satoh M: **Differential patterns of c-fos mRNA expression in the amygdaloid nuclei induced by chemical somatic and visceral noxious stimuli in rats.** *Neurosci Lett* 2003, **344**:197–200.
25. Suwanprathes P, Ngu M, Ing A, Hunt G, Seow F: **c-Fos immunoreactivity in the brain after esophageal acid stimulation.** *Am J Med* 2003, **115**(Suppl 3A):315–385.
26. Greenwood-Van Meerveld B, Johnson AC, Schulkin J, Myers DA: **Long-term expression of corticotropin-releasing factor (CRF) in the paraventricular nucleus of the hypothalamus in response to an acute colonic inflammation.** *Brain Res* 2006, **1071**:91–96.
27. Han JS, Neugebauer V: **Synaptic plasticity in the amygdala in a visceral pain model in rats.** *Neurosci Lett* 2004, **361**:254–257.
28. Bon K, Lanteri-Minet M, Michiels JF, Menetrey D: **Cyclophosphamide cystitis as a model of visceral pain in rats: a c-fos and Krox-24 study at telencephalic levels, with a note on pituitary adenylate cyclase activating polypeptide (PACAP).** *Exp Brain Res* 1998, **122**:165–174.
29. Nishii H, Nomura M, Aono H, Fujimoto N, Matsumoto T: **Up-regulation of galanin and corticotropin-releasing hormone mRNAs in the key hypothalamic and amygdaloid nuclei in a mouse model of visceral pain.** *Regul Pept* 2007, **141**:105–112.
30. Carrasquillo Y, Gereau RW: **Activation of the extracellular signal-regulated kinase in the amygdala modulates pain perception.** *J Neurosci* 2007, **27**:1543–1551.
31. Carrasquillo Y, Gereau RW: **Hemispheric lateralization of a molecular signal for pain modulation in the amygdala.** *Mol Pain* 2008, **4**:24.
32. Cheng SJ, Chen CC, Yang HW, Chang YT, Bai SW, Yen CT, Min MY: **Role of extracellular signal-regulated kinase in synaptic transmission and plasticity of a nociceptive input on capsular central amygdaloid neurons in normal and acid-induced muscle pain mice.** *J Neurosci* 2011, **31**:2258–2270.
33. Neugebauer V, Li W: **Differential sensitization of amygdala neurons to afferent inputs in a model of arthritic pain.** *J Neurophysiol* 2003, **89**:716–727.
34. Ji G, Neugebauer V: **Hemispheric lateralization of pain processing by amygdala neurons.** *J Neurophysiol* 2009, **102**:2253–2264.
35. Neugebauer V, Li W, Bird GC, Bhawe G, Gereau RW: **Synaptic plasticity in the amygdala in a model of arthritic pain: differential roles of metabotropic glutamate receptors 1 and 5.** *J Neurosci* 2003, **23**:52–63.
36. Bird GC, Lash LL, Han JS, Zou X, Willis WD, Neugebauer V: **Protein kinase A-dependent enhanced NMDA receptor function in pain-related synaptic plasticity in rat amygdala neurons.** *J Physiol* 2005, **564**:907–921.
37. Goncalves L, Dickenson AH: **Asymmetric time-dependent activation of right central amygdala neurons in rats with peripheral neuropathy and pregabalin modulation.** *Eur J Neurosci* 2012, **36**:3204–3213.
38. Ikeda R, Takahashi Y, Inoue K, Kato F: **NMDA receptor-independent synaptic plasticity in the central amygdala in the rat model of neuropathic pain.** *Pain* 2007, **127**:161–172.
39. Ulrich-Lai YM, Xie W, Meij JT, Dolgas CM, Yu L, Herman JP: **Limbic and HPA axis function in an animal model of chronic neuropathic pain.** *Physiol Behav* 2006, **88**:67–76.
40. Rouwette T, Vanelderen P, de Reus M, Loohuis NO, Giele J, van Egmond J, Scheenen W, Scheffer GJ, Roubos E, Vissers K, Kozicz T: **Experimental neuropathy increases limbic forebrain CRF.** *Eur J Pain* 2012, **16**:61–71.
41. Goncalves L, Silva R, Pinto-Ribeiro F, Pego JM, Bessa JM, Pertovaara A, Sousa N, Almeida A: **Neuropathic pain is associated with depressive behaviour and induces neuroplasticity in the amygdala of the rat.** *Exp Neurol* 2008, **213**:48–56.
42. Traub RJ, Silva E, Gebhart GF, Solodkin A: **Noxious colorectal distention induced-c-Fos protein in limbic brain structures in the rat.** *Neurosci Lett* 1996, **215**:165–168.
43. Bornhovd K, Quante M, Glauche V, Bromm B, Weiller C, Buchel C: **Painful stimuli evoke different stimulus-response functions in the amygdala, prefrontal, insula and somatosensory cortex: a single-trial fMRI study.** *Brain* 2002, **125**:1326–1336.
44. Petrovic P, Carlsson K, Petersson KM, Hansson P, Ingvar M: **Context-dependent deactivation of the amygdala during pain.** *J Cogn Neurosci* 2004, **16**:1289–1301.
45. Bonaz B, Baciou M, Papillon E, Bost R, Gueddah N, Le Bas JF, Fournet J, Segebarth C: **Central processing of rectal pain in patients with irritable bowel syndrome: an fMRI study.** *Am J Gastroenterol* 2002, **97**:654–661.
46. Veinante P, Freund-Mercier MJ: **Branching patterns of central amygdaloid nucleus efferents in the Rat: single-axon reconstructions.** *Ann N Y Acad Sci* 2003, **985**:552–553.
47. Pavlovic ZW, Bodnar RJ: **Opioid supraspinal analgesic synergy between the amygdala and periaqueductal gray in rats.** *Brain Res* 1998, **779**:158–169.
48. Hermann DM, Luppi PH, Peyron C, Hinckel P, Jouvett M: **Afferent projections to the rat nuclei raphe magnus, raphe pallidus and reticularis gigantocellularis pars alpha demonstrated by iontophoretic application of cholera toxin (subunit b).** *J Chem Neuroanat* 1997, **13**:1–21.
49. Almeida A, Cobos A, Tavares I, Lima D: **Brain afferents to the medullary dorsal reticular nucleus: a retrograde and anterograde tracing study in the rat.** *Eur J Neurosci* 2002, **16**:81–95.
50. Jin X, Cui N, Zhong W, Jin XT, Jiang C: **GABA-ergic synaptic inputs of locus coeruleus neurons in wild-type and MeCP2-null mice.** *Am J Physiol Cell Physiol* 2013. doi:10.1152/ajpcell.00399.2012. in press.
51. Zahm DS, Cheng AY, Lee TJ, Ghobadi CW, Schwartz ZM, Geisler S, Parsely KP, Gruber C, Veh RW: **Inputs to the midbrain dopaminergic complex in the rat, with emphasis on extended amygdala-recipient sectors.** *J Comp Neurol* 2011, **519**:3159–3188.
52. Wallace DM, Magnuson DJ, Gray TS: **Organization of amygdaloid projections to brainstem dopaminergic, noradrenergic, and adrenergic cell groups in the rat.** *Brain Res Bull* 1992, **28**:447–454.
53. Barrot M: **Tests and models of nociception and pain in rodents.** *Neuroscience* 2012, **211**:39–50.
54. Calvino B, Levesque G, Besson JM: **Possible involvement of the amygdaloid complex in morphine analgesia as studied by electrolytic lesions in rats.** *Brain Res* 1982, **233**:221–226.
55. Helmstetter FJ, Bellgowan PS: **Lesions of the amygdala block conditional hypoaesthesia on the tail flick test.** *Brain Res* 1993, **612**:253–257.
56. Manning BH, Mayer DJ: **The central nucleus of the amygdala contributes to the production of morphine antinociception in the rat tail-flick test.** *J Neurosci* 1995, **15**:8199–8213.
57. Han Y, Yu LC: **Involvement of oxytocin and its receptor in nociceptive modulation in the central nucleus of amygdala of rats.** *Neurosci Lett* 2009, **454**:101–104.

58. Cui XY, Lundeberg T, Yu LC: Role of corticotropin-releasing factor and its receptor in nociceptive modulation in the central nucleus of amygdala in rats. *Brain Res* 2004, **995**:23–28.
59. Xu W, Lundeberg T, Wang YT, Li Y, Yu LC: Antinociceptive effect of calcitonin gene-related peptide in the central nucleus of amygdala: activating opioid receptors through amygdala-periaqueductal gray pathway. *Neuroscience* 2003, **118**:1015–1022.
60. Jin WY, Liu Z, Liu D, Yu LC: Antinociceptive effects of galanin in the central nucleus of amygdala of rats, an involvement of opioid receptors. *Brain Res* 2010, **1320**:16–21.
61. Ahn DK, Kim KH, Ju JS, Kwon S, Park JS: Microinjection of arginine vasopressin into the central nucleus of amygdala suppressed nociceptive jaw opening reflex in freely moving rats. *Brain Res Bull* 2001, **55**:117–121.
62. Kalivas PW, Gau BA, Nemeroff CB, Prange AJ Jr: Antinociception after microinjection of neurotensin into the central amygdaloid nucleus of the rat. *Brain Res* 1982, **243**:279–286.
63. Ahn DK, Kim YS, Park JS: Central-amygdaloid carbachol suppressed nociceptive jaw opening reflex in freely moving rats. *Prog Neuropsychopharmacol Biol Psychiatry* 1999, **23**:685–695.
64. Ortiz JP, Heinricher MM, Selden NR: Noradrenergic agonist administration into the central nucleus of the amygdala increases the tail-flick latency in lightly anesthetized rats. *Neuroscience* 2007, **148**:737–743.
65. Oliveira MA, Prado WA: Antinociception and behavioral manifestations induced by intracerebroventricular or intra-amygdaloid administration of cholinergic agonists in the rat. *Pain* 1994, **57**:383–391.
66. Leite-Panissi CR, Brentegani MR, Menescal-de-Oliveira L: Cholinergic-opioidergic interaction in the central amygdala induces antinociception in the guinea pig. *Braz J Med Biol Res* 2004, **37**:1571–1579.
67. Manning BH, Martin WJ, Meng JD: The rodent amygdala contributes to the production of cannabinoid-induced antinociception. *Neuroscience* 2003, **120**:1157–1170.
68. Sarhan M, Pawlowski SA, Barthas F, Yalcin I, Kaufling J, Dardente H, Zachariou V, Dileone RJ, Barrot M, Veinante P: BDNF parabrachio-amygdaloid pathway in morphine-induced analgesia. *Int J Neuropsychopharmacol* 2013. doi:10.1017/S146114571200168X. in press.
69. Bellgowan PS, Helmstetter FJ: Neural systems for the expression of hypoalgesia during nonassociative fear. *Behav Neurosci* 1996, **110**:727–736.
70. Helmstetter FJ: The amygdala is essential for the expression of conditional hypoalgesia. *Behav Neurosci* 1992, **106**:518–528.
71. Manning BH, Mayer DJ: The central nucleus of the amygdala contributes to the production of morphine antinociception in the formalin test. *Pain* 1995, **63**:141–152.
72. Tanimoto S, Nakagawa T, Yamauchi Y, Minami M, Satoh M: Differential contributions of the basolateral and central nuclei of the amygdala in the negative affective component of chemical somatic and visceral pains in rats. *Eur J Neurosci* 2003, **18**:2343–2350.
73. Gao YJ, Ren WH, Zhang YQ, Zhao ZQ: Contributions of the anterior cingulate cortex and amygdala to pain- and fear-conditioned place avoidance in rats. *Pain* 2004, **110**:343–353.
74. Pedersen LH, Scheel-Kruger J, Blackburn-Munro G: Amygdala GABA-A receptor involvement in mediating sensory-discriminative and affective-motivational pain responses in a rat model of peripheral nerve injury. *Pain* 2007, **127**:17–26.
75. Ansah OB, Bourbia N, Goncalves L, Almeida A, Pertovaara A: Influence of amygdaloid glutamatergic receptors on sensory and emotional pain-related behavior in the neuropathic rat. *Behav Brain Res* 2010, **209**:174–178.
76. Crock LW, Kolber BJ, Morgan CD, Sadler KE, Vogt SK, Bruchas MR, Gereau RW: Central amygdala metabotropic glutamate receptor 5 in the modulation of visceral pain. *J Neurosci* 2012, **32**:14217–14226.
77. Kolber BJ, Montana MC, Carrasquillo Y, Xu J, Heinemann SF, Muglia LJ, Gereau RW: Activation of metabotropic glutamate receptor 5 in the amygdala modulates pain-like behavior. *J Neurosci* 2010, **30**:8203–8213.
78. Han JS, Neugebauer V: mGluR1 and mGluR5 antagonists in the amygdala inhibit different components of audible and ultrasonic vocalizations in a model of arthritic pain. *Pain* 2005, **113**:211–222.
79. Palazzo E, Fu Y, Ji G, Maione S, Neugebauer V: Group III mGluR7 and mGluR8 in the amygdala differentially modulate nocifensive and affective pain behaviors. *Neuropharmacology* 2008, **55**:537–545.
80. Ortiz JP, Close LN, Heinricher MM, Selden NR: Alpha(2)-noradrenergic antagonist administration into the central nucleus of the amygdala blocks stress-induced hypoalgesia in awake behaving rats. *Neuroscience* 2008, **157**:223–228.
81. Deyama S, Takishita A, Tanimoto S, Ide S, Nakagawa T, Satoh M, Minami M: Roles of beta- and alpha2-adrenoceptors within the central nucleus of the amygdala in the visceral pain-induced aversion in rats. *J Pharmacol Sci* 2010, **114**:123–126.
82. Han JS, Adwanikar H, Li Z, Ji G, Neugebauer V: Facilitation of synaptic transmission and pain responses by CGRP in the amygdala of normal rats. *Mol Pain* 2010, **6**:10.
83. Han JS, Li W, Neugebauer V: Critical role of calcitonin gene-related peptide 1 receptors in the amygdala in synaptic plasticity and pain behavior. *J Neurosci* 2005, **25**:10717–10728.
84. Ji G, Fu Y, Adwanikar H, Neugebauer V: Non-pain-related CRF1 activation in the amygdala facilitates synaptic transmission and pain responses. *Mol Pain* 2013, **9**:2.
85. Ji G, Fu Y, Ruppert KA, Neugebauer V: Pain-related anxiety-like behavior requires CRF1 receptors in the amygdala. *Mol Pain* 2007, **3**:13.
86. Fu Y, Neugebauer V: Differential mechanisms of CRF1 and CRF2 receptor functions in the amygdala in pain-related synaptic facilitation and behavior. *J Neurosci* 2008, **28**:3861–3876.
87. Qin C, Greenwood-Van Meerveld B, Foreman RD: Spinal neuronal responses to urinary bladder stimulation in rats with corticosterone or aldosterone onto the amygdala. *J Neurophysiol* 2003, **90**:2180–2189.
88. Qin C, Greenwood-Van Meerveld B, Myers DA, Foreman RD: Corticosterone acts directly at the amygdala to alter spinal neuronal activity in response to colorectal distension. *J Neurophysiol* 2003, **89**:1343–1352.
89. Myers B, Dittmeyer K, Greenwood-Van Meerveld B: Involvement of amygdaloid corticosterone in altered visceral and somatic sensation. *Behav Brain Res* 2007, **181**:163–167.
90. Myers B, Greenwood-Van Meerveld B: Corticosteroid receptor-mediated mechanisms in the amygdala regulate anxiety and colonic sensitivity. *Am J Physiol Gastrointest Liver Physiol* 2007, **292**:G1622–G1629.
91. Fu Y, Han J, Ishola T, Scerbo M, Adwanikar H, Ramsey C, Neugebauer V: PKA and ERK, but not PKC, in the amygdala contribute to pain-related synaptic plasticity and behavior. *Mol Pain* 2008, **4**:26.
92. Crown ED, King TE, Meagher MW, Grau JW: Shock-induced hyperalgesia: III. Role of the bed nucleus of the stria terminalis and amygdaloid nuclei. *Behav Neurosci* 2000, **114**:561–573.
93. Rhudy JL, Meagher MW: Negative affect: effects on an evaluative measure of human pain. *Pain* 2003, **104**:617–626.
94. Yalcin I, Bohren Y, Waltisperger E, Sage-Ciocca D, Yin JC, Freund-Mercier MJ, Barrot M: A time-dependent history of mood disorders in a murine model of neuropathic pain. *Biol Psychiatry* 2011, **70**:946–953.
95. Calvino B, Besson JM, Boehrer A, Depaulis A: Ultrasonic vocalization (22–28 kHz) in a model of chronic pain, the arthritic rat: effects of analgesic drugs. *NeuroReport* 1996, **7**:581–584.
96. Kulkarni B, Bentley DE, Elliott R, Julyan PJ, Boger E, Watson A, Boyle Y, El-Derey W, Jones AK: Arthritic pain is processed in brain areas concerned with emotions and fear. *Arthritis Rheum* 2007, **56**:1345–1354.
97. Petrovic P, Ingvar M, Stone-Elander S, Petersson KM, Hansson P: A PET activation study of dynamic mechanical allodynia in patients with mononeuropathy. *Pain* 1999, **83**:459–470.
98. Yoshino A, Okamoto Y, Onoda K, Yoshimura S, Kunisato Y, Demoto Y, Okada G, Yamawaki S: Sadness enhances the experience of pain via neural activation in the anterior cingulate cortex and amygdala: an fMRI study. *NeuroImage* 2010, **50**:1194–1201.
99. Younger J, Aron A, Parke S, Chatterjee N, Mackey S: Viewing pictures of a romantic partner reduces experimental pain: involvement of neural reward systems. *PLoS One* 2010, **5**:e13309.
100. Delaney AJ, Crane JW, Sah P: Noradrenaline modulates transmission at a central synapse by a presynaptic mechanism. *Neuron* 2007, **56**:880–892.
101. Yamano M, Hillyard CJ, Girgis S, Emson PC, MacIntyre I, Tohyama M: Projection of neurotensin-like immunoreactive neurons from the lateral parabrachial area to the central amygdaloid nucleus of the rat with reference to the coexistence with calcitonin gene-related peptide. *Exp Brain Res* 1988, **71**:603–610.
102. Salio C, Averill S, Priestley JV, Merighi A: Costorage of BDNF and neuropeptides within individual dense-core vesicles in central and peripheral neurons. *Dev Neurobiol* 2007, **67**:326–338.

103. Sun N, Cassel MD: Intrinsic GABAergic neurons in the rat central extended amygdala. *J Comp Neurol* 1993, **330**:381–404.
104. Ciochi S, Herry C, Grenier F, Wolff SB, Letzkus JJ, Vlachos I, Ehrlich I, Sprengel R, Deisseroth K, Stadler MB, et al: Encoding of conditioned fear in central amygdala inhibitory circuits. *Nature* 2010, **468**:277–282.
105. Haubensak W, Kunwar PS, Cai H, Ciochi S, Wall NR, Ponnusamy R, Biag J, Dong HW, Deisseroth K, Callaway EM, et al: Genetic dissection of an amygdala microcircuit that gates conditioned fear. *Nature* 2010, **468**:270–276.
106. Dong YL, Fukazawa Y, Wang W, Kamasawa N, Shigemoto R: Differential postsynaptic compartments in the laterocapsular division of the central nucleus of amygdala for afferents from the parabrachial nucleus and the basolateral nucleus in the rat. *J Comp Neurol* 2010, **518**:4771–4791.
107. Li W, Neugebauer V: Differential roles of mGluR1 and mGluR5 in brief and prolonged nociceptive processing in central amygdala neurons. *J Neurophysiol* 2004, **91**:13–24.
108. Ren W, Neugebauer V: Pain-related increase of excitatory transmission and decrease of inhibitory transmission in the central nucleus of the amygdala are mediated by mGluR1. *Mol Pain* 2010, **6**:93.
109. Han JS, Bird GC, Neugebauer V: Enhanced group III mGluR-mediated inhibition of pain-related synaptic plasticity in the amygdala. *Neuropharmacology* 2004, **46**:918–926.
110. Han JS, Fu Y, Bird GC, Neugebauer V: Enhanced group II mGluR-mediated inhibition of pain-related synaptic plasticity in the amygdala. *Mol Pain* 2006, **2**:18.
111. Li W, Neugebauer V: Differential changes of group II and group III mGluR function in central amygdala neurons in a model of arthritic pain. *J Neurophysiol* 2006, **96**:1803–1815.
112. Li Z, Ji G, Neugebauer V: Mitochondrial reactive oxygen species are activated by mGluR5 through IP3 and activate ERK and PKA to increase excitability of amygdala neurons and pain behavior. *J Neurosci* 2011, **31**:1114–1127.
113. Li W, Neugebauer V: Block of NMDA and non-NMDA receptor activation results in reduced background and evoked activity of central amygdala neurons in a model of arthritic pain. *Pain* 2004, **110**:112–122.
114. Jaferi A, Pickel VM: Mu-opioid and corticotropin-releasing-factor receptors show largely postsynaptic co-expression, and separate presynaptic distributions, in the mouse central amygdala and bed nucleus of the stria terminalis. *Neuroscience* 2009, **159**:526–539.
115. Zhu W, Pan ZZ: Mu-opioid-mediated inhibition of glutamate synaptic transmission in rat central amygdala neurons. *Neuroscience* 2005, **133**:97–103.
116. Chieng BC, Christie MJ, Osborne PB: Characterization of neurons in the rat central nucleus of the amygdala: cellular physiology, morphology, and opioid sensitivity. *J Comp Neurol* 2006, **497**:910–927.
117. Narita M, Kaneko C, Miyoshi K, Nagumo Y, Kuzumaki N, Nakajima M, Nanjo K, Matsuzawa K, Yamazaki M, Suzuki T: Chronic pain induces anxiety with concomitant changes in opioidergic function in the amygdala. *Neuropsychopharmacology* 2006, **31**:739–750.
118. Finnegan TF, Chen SR, Pan HL: Effect of the mu opioid on excitatory and inhibitory synaptic inputs to periaqueductal gray-projecting neurons in the amygdala. *J Pharmacol Exp Ther* 2005, **312**:441–448.
119. Deyama S, Nakagawa T, Kaneko S, Uehara T, Minami M: Involvement of the bed nucleus of the stria terminalis in the negative affective component of visceral and somatic pain in rats. *Behav Brain Res* 2007, **176**:367–371.
120. Deyama S, Katayama T, Kondoh N, Nakagawa T, Kaneko S, Yamaguchi T, Yoshioka M, Minami M: Role of enhanced noradrenergic transmission within the ventral bed nucleus of the stria terminalis in visceral pain-induced aversion in rats. *Behav Brain Res* 2009, **197**:279–283.
121. Deyama S, Katayama T, Ohno A, Nakagawa T, Kaneko S, Yamaguchi T, Yoshioka M, Minami M: Activation of the beta-adrenoceptor-protein kinase A signaling pathway within the ventral bed nucleus of the stria terminalis mediates the negative affective component of pain in rats. *J Neurosci* 2008, **28**:7728–7736.
122. Deyama S, Ide S, Kondoh N, Yamaguchi T, Yoshioka M, Minami M: Inhibition of noradrenaline release by clonidine in the ventral bed nucleus of the stria terminalis attenuates pain-induced aversion in rats. *Neuropharmacology* 2011, **61**:156–160.
123. Ide S, Hara T, Ohno A, Tamano R, Koseki K, Naka T, Maruyama C, Kaneda K, Yoshioka M, Minami M: Opposing roles of corticotropin-releasing factor and neuropeptide Y within the dorsolateral bed nucleus of the stria terminalis in the negative affective component of pain in rats. *J Neurosci* 2013, **33**:5881–5894.
124. Connell K, Bolton N, Olsen D, Piomelli D, Hohmann AG: Role of the basolateral nucleus of the amygdala in endocannabinoid-mediated stress-induced analgesia. *Neurosci Lett* 2006, **397**:180–184.
125. Rea K, Olango WM, Harhen B, Kerr DM, Galligan R, Fitzgerald S, Moore M, Roche M, Finn DP: Evidence for a role of GABAergic and glutamatergic signalling in the basolateral amygdala in endocannabinoid-mediated fear-conditioned analgesia in rats. *Pain* 2013, **154**:576–585.
126. Rea K, Roche M, Finn DP: Modulation of conditioned fear, fear-conditioned analgesia, and brain regional c-Fos expression following administration of muscimol into the rat basolateral amygdala. *J Pain* 2011, **12**:712–721.
127. Helmstetter FJ: Stress-induced hypoalgesia and defensive freezing are attenuated by application of diazepam to the amygdala. *Pharmacol Biochem Behav* 1993, **44**:433–438.
128. Nandigama P, Borszcz GS: Affective analgesia following the administration of morphine into the amygdala of rats. *Brain Res* 2003, **959**:343–354.
129. Helmstetter FJ, Bellgowan PS, Poore LH: Microinfusion of mu but not delta or kappa opioid agonists into the basolateral amygdala results in inhibition of the tail flick reflex in pentobarbital-anesthetized rats. *J Pharmacol Exp Ther* 1995, **275**:381–388.
130. McGaraughty S, Heinricher MM: Microinjection of morphine into various amygdaloid nuclei differentially affects nociceptive responsiveness and RVM neuronal activity. *Pain* 2002, **96**:153–162.
131. Chen J, Song Y, Yang J, Zhang Y, Zhao P, Zhu XJ, Su HC: The contribution of TNF-alpha in the amygdala to anxiety in mice with persistent inflammatory pain. *Neurosci Lett* 2013, **541**:275–280.
132. Ji G, Sun H, Fu Y, Li Z, Pais-Vieira M, Galhardo V, Neugebauer V: Cognitive impairment in pain through amygdala-driven prefrontal cortical deactivation. *J Neurosci* 2010, **30**:5451–5464.
133. Deyama S, Yamamoto J, Machida T, Tanimoto S, Nakagawa T, Kaneko S, Satoh M, Minami M: Inhibition of glutamatergic transmission by morphine in the basolateral amygdaloid nucleus reduces pain-induced aversion. *Neurosci Res* 2007, **59**:199–204.
134. Watabe AM, Ochiai T, Nagase M, Takahashi Y, Sato M, Kato F: Synaptic potentiation in the nociceptive amygdala following fear learning in mice. *Mol Brain* 2013, **6**:11.

doi:10.1186/2049-9256-1-9

Cite this article as: Veinante et al.: The amygdala between sensation and affect: a role in pain. *Journal of Molecular Psychiatry* 2013 **1**:9.

Submit your next manuscript to BioMed Central and take full advantage of:

- Convenient online submission
- Thorough peer review
- No space constraints or color figure charges
- Immediate publication on acceptance
- Inclusion in PubMed, CAS, Scopus and Google Scholar
- Research which is freely available for redistribution

Submit your manuscript at
www.biomedcentral.com/submit

